

Impact of Subclinical Endometritis on Some Fertility Traits of Dairy Cows

Constantin Găvan¹, Cosmin Şonea²

¹Agriculture Research and Development Station Şimnic, Craiova, Şoseau Bălceşti, no. 54, 200721, Dolj, Romania.

²University of Agronomical Sciences and Veterinary Medicine Bucharest, Romania

Abstract

The aim of this retrospective study was to determine that most suitable Polymorphonuclear leukocytes (PMNs) threshold for diagnosis of subclinical endometritis (SE), the causes of this disorder and its effect on subsequent reproductive parameters of Holstein Friesian cows. Uterine cytology was executed on 140 Holstein cows at 28-30 days postpartum (pp) to calculate the PMN cells. A threshold of 16% PMNs above which some of the reproductive parameters were significantly affected was used. The retained fetal membranes (Odds ratio; OR = 3.02) and metritis (OR = 4.92) were the causes for SE. subclinical endometritis and metabolic disorders (milk fever and ketosis) affected the resumption of pp cyclicity at 28-30 days in milk. Cows with SE were less likely to conceive after their artificial insemination (OR = 0.44 p<0.05) than cows without S.E., and needed more artificial inseminations to conceive (2.4 vs. 2.1.). A PMN threshold of 16% was good enough to detect SE at 28-30 days pp. the RFMs and metritis were causal factors for SE.

Keywords: cytological endometritis, subclinical endometritis, polymorphonuclear cells, reproductive parameters, causal factors.

1. Introduction

Subclinical endometritis (SE) described as an inflammation of the endometrium without clinical signs and often without evidence of infection [1-3]. This uterine disease has been associated with in later time in restarting ovarian activity postpartum, extended intervals from parturition to first AI service, more days open, low conception rates, and high culling rates [4-6].

Subclinical endometritis has also been reported as “cytological endometritis” [7, 8] on the basis of “an elevated ratio of polymorphonuclear cells (PMN)” in endometrial cytology samples. In their report, Dubuc et al., (2010) [8], indicated that cytological endometritis and clinical endometritis may actually represent different manifestations of

reproductive tract disease and inflammation, and not a difference in the gravity of the same disease. The uterine pathogens affect reproduction by direct endometrial injury and by generating endotoxins [9]. The bacterial endotoxins have some effects on cow reproduction [10-12]. But, in the case SE, some authors reported that bacterial populations presented in the uterus of cows with SE did not differ from those presented in the healthy uterus of dairy cows [1, 3].

Some previous reports demonstrated negative impacts of elevated concentrations of non-esterified fatty acids (NEFA), beta-hydroxybutyric acid (BHBA), bilirubin, and urea on the function of PMN [13, 14]. Also, in some past reports [15-19] showed that cows with SE had an increased endometrial mRNA expression and elevated serum concentration of pro-inflammatory mediators as compared with healthy cows.

In addition, the type of fatty acids present in cow's ration can affect cellular immune function [20].

* Corresponding author: C. Găvan,
scda_simnic@yahoo.com

The diagnosis of SE can be due by endometrial cytology [19]. Also, ultrasonography was used, but in some reports ultrasonography method was less sensitive than endometrial cytology [19, 21, 22].

Different thresholds of PMN for diagnosis of SE in cows were accepted [23]. The cutoff PMN% reported has varied between 4 and 25% and has depending on the postpartum period at which the diagnostic was done (18% OMN at 20-33 days postpartum (pp); 10% PMN at 34-47 days pp and 5% PMN between 21-66 days pp [24].

The prevalence of SE reported in the literature in the literature in general, decreases with increasing time pp. Many affected cows recover spontaneously [25-27], and this can happen to four [28] to eight [29] weeks pp as a result pf the estrus. Circulating estradiol concentrations at estrus stimulate uterine motility and PMN function and this is an aid for eliminating abnormal fluid and get control of inflammatory changes [30].

Regarding the effects of SE on reproductive performance some authors [31, 32] did not reported significant effects on reproduction and many others reported a variety of negative effects on fertility [19, 7, 21, 8, 33, 34, 35, 32]. Affected parameters were: Days Open, Pregnancy rate, Postpartum anestrus, First-service pregnancy rate, Services per conception, transferable embryo recovery rate, culling rate, follicle development and ovulation, pregnancy rate at 150 and 250 days pp.

Also, clinical and subclinical endometritis have been associated with a declined milk production [36].

We hypothesized that the PMN threshold used to define SE and the timing of evaluation during pp period vary among herds and the choices of optimal PMN threshold should be calculated for each herd. The aim of this study was to determine the most suitable PMN threshold for diagnosis of SE, the risk factors and its effect on subsequent reproductive parameters of Holstein Friesian cows.

2. Materials and methods

2.1. Animals

Animals involved in this study were from Agricultural Research and Development Station (ARDS) Șimnic-Craiova (182 m above sea level

44°19' N, 23°48' E). The dairy farm of ARDS Șimnic-Craiova-Romania has a herd size of 280 Holstein Friesian dairy animals, and milk production of the cows was on average 9440 kg per 305 days of lactation.

All the cows were managed in loose housing with access to dry lot prepartum and postpartum. The cows are milked twice daily at 12 hours interval (morning and evening) in a DeLaval milking parlour.

Cows and heifers are fed in the morning and afternoon with diets formulated to meet or exceed nutritional requirements for dry and lactating dairy cows. The diets consist of high energy ration based on a grain by-products, grain mix and home-grown forages.

Additionally, the cows' heifers have access to pasture (alfalfa or ryegrass). A prepartum and a postpartum diet is presented in table 1.

Table 1. Prepartum and postpartum diet (dry mother basis)

Item	Prepartum %	Postpartum %
A. Ingredients		
Alfalfa hay	26	29
Grass hay	45	5.5
Corn silage	10	16
Grain mix	16	47
Mineral mix	3	2.5
Total	100	100
B. Nutrient profile		
Net energy lactation	1.28	1.6
Crude protein	12	19
Neutral detergent fiber	46	28
Acid detergent fiber	40	20
Starch	14	27
Calcium	1.4	1.3
Phosphorus	0.3	0.4
Magnesium	0.4	0.4
Kalium	1.5	1.3
Natrium	0.3	0.4
Chloride	1.1	0.4
Sulf	0.4	0.2
DCAD mEq/100 of DM*	5	26.9

*Calculated as DCAD = (mEq of Na + mEq of K) – (mEq of S + mEq of Cl)

Pregnant cows are dried-off 50 ± 10 days before the expected calving date. In the prepartum pen cows are closely monitored by the farm or research personnel for signs of parturition. In the parturition pen all events are recorded.

The experiment was performed in compliance with the European Union Directive 86/609 Ec. and National legislation.

For this study, a total of 162 Holstein Friesian cows' parity 1-6 were enrolled. All this cows with calvings between 1st January 2021 and December 2022 were evaluated for uterine health status between 28 to 30 days in milk (DIM), using rectal palpation, vaginoscopy or transrectal ultrasonography (US). Cows in estrus were not used. Cows with signs of abnormal discharge of uterine fluid, abnormal fluid into the uterus or pathologic enlargement of the uterus were recorded as having clinical endometritis (n=22). Cow with no signs of endometritis were sampled for uterine cytology (n=140). Uterine cytology was performed using a cytobrush modified for use in large animals [37]. On short: The handle of the cytobrush is cut to 3 cm in length, threaded onto a stainless-steel rod (65 cm length), and placed in a stainless-steel tube (5 mm in diameter and 50 cm length). The instrument is placed in a sanitary plastic sleeve for protection. The vulva is cleaned and the lubricated instrument is passed through vagina to external cervical OS; the sanitary plastic sleeve is punctured and the instrument is advanced through the cervix into the base of the larger horn, and at this point the stainless-steel tube is retracted to expose the cytobrush. The sample is collected by rotating the cytobrush (clockwise direction) on the uterine wall. Then the cytobrush is retracted into tube, and removed from the uterus.

The instrument can be steam sterilized for 4 minutes between uses. By rolling the cytobrush onto a clean microscope glass slide and allowed to air-dry, all slides are stained. We used a Diff-Quick stain kit, according to the manufacturer's instructions. All slides were examined under microscope (400 X magnification). Up to 200 endometrial cells and PMN were counted and the percentage of PMNs present was calculated per each slide.

The health disorders used in this study were: Calving difficulty, Retained fetal membranes, Septicemic metritis, Ketosis and milk fever. The veterinary waiting period from calving to first artificial insemination (AI) was 50 days. All cows were AI at detected estrus. Pregnancy diagnosis was made at 45-50 days after AI, based on

transrectal palpation. Reproductive parameters were recorded up to 180 days pp or until pregnancy.

2.2. Study design

Calving difficulty was assessed according to the degree of assistance (1 = no assistance, 2 = easy assistance, 3 = using some force, 4 = using significant force and 5 = caesarian section). Cows with difficulty scores >2 were categorized as having dystocia [38].

Retained fetal membranes was recorded as retention for longer than 12 hours after calving. Metritis was recorded as watery, fetid uterine discharge during 10 days pp and fever ($\geq 39.5^{\circ}\text{C}$) ketosis was recorded using clinical signs: depression, anorexia, and odor of acetone on the breath. Milk fever was recorded as recumbency after calving. The research personnel examined ovarian structures follicles, corpus luteum, and uterus by transrectal palpation and ultrasonography. Resumption of cyclicity within 28-30 days pp was recorded by detection of corpus luteum. First, we calculated the % PMN threshold, then prevalence of SE and casual factors for SE.

2.3. Statistical analysis

The data were entered into M. Excel computer program 2007.

Stata version 14 was used to express the results. The causal factors for SE and probabilities of resumption cyclicity, and conception after AI were calculated using logistic regression. Odds ratio, 95% Confidence Interval and p value were calculated using logistic regression. A p value ≤ 0.05 was considered statistically significant.

3. Results and discussion

The threshold for PMN proportion, above which some of the reproductive parameters were significantly affected was 16%. Using this threshold at 28-30 days after parturition we described SE.

The prevalence of SE was 34.3% (48 from 140 cows). Some of the risk factors for SE are presented in table 2.

Table 2. Odds ratios (OR) of the risk factors for subclinical endometritis

Variables	% of SE incidence (Number of cows)	Odds ratio	95% CI	p value
Retained fetal membranes				
No	15.21 (14/92)	Reference	1.44-6.34	0.00176
Yes	54.16 (26/48)	3.020		
Metritis				
No	10.87 (10/92)	Reference	2.043-11.848	0.0004
Yes	37.5 (18/48)	4.920		
Dystocia				
No	13.04 (12/92)	Reference	0.3337-2.7182	0.9273
Yes	12.5 (6/48)	0.9524		
Metabolic disorders*				
No	10.87 (10/92)	Reference	0.221 to 2.5150	0.6359
Yes	8.33 (4/48)	0.7455		

SU = subclinical endometritis

CI = confidence interval

* = metabolic disorders consist of ketosis and milk fever.

The presence of retained fetal membranes (RFM) in cows after parturition and metritis during first 10 days postpartum significantly affected the incidence of SE (table 2). No effect for dystocia and metritis during first 10 days postpartum significantly affected the incidence of SE (table

2). No effect for dystocia and metabolic disorder ($p > 0.1$). Cows with RFM (OR = 3.02) and with metritis during 10 days postpartum (OR = 4.92) had a higher risk of SE than cows without SE (table 2).

Table 3. The probability of resumption of cyclicity within 28-30 days in milk.

Variables	% resumption rate (No. of cows)	Odds ratio	95% CI	p value
SE				
No	47.82 (44/92)	Reference		
Yes	20.83 (10/48)	0.3133	0.1396-0.7031	0.0049
RFM				
No	65 (65/100)	Reference		
Yes	50 (20/40)	0.5385	0.2560-1.1327	0.10
Metritis				
Yes	58.03 (65/112)	Reference		
No	71.42 (20/28)	1.8077	0.7337-4.4539	0.1981
Dystocia				
No	57.4 (70/122)	Reference		
Yes	55.55(10/18)	0.9286	0.3428-2.5154	0.8841
Metabolic disorders				
No	53.96 (68/126)	Reference		
Yes	71.42 (10/14)	4.6324	1.400-15.325	0.0120

SE = Subclinical endometritis;

RFM = Retained fetal membranes;

CI = Confidence interval.

Subclinical endometritis and metabolic disorders affected the resumption of postpartum cyclicity at 28-30 days in milk. Cows with SE had a decrease

probability (OR = 0.31, $P < 0.05$) of resumption of cyclicity after parturition than cows without SE (table 3).

Table 4. The probability of conception after the first insemination.

Variable	% conception rate at first insemination	Odds ratio	95% CI	p value
SE	No 45.65 (42/92)	Reference		
	Yes 27.1 (13/48)	0.4422	0.2073-0.9430	0.0347
RFM	No 46 (46/100)	Reference		
	Yes 30 (10/40)	0.3913	0.1729-0.8855	0.0243
Metritis	Yes 44.64 (50/112)	Reference		
	No 28.57 (6/28)	0.3382	0.1274-0.8980	0.0296
Dystocia	No 49.2 (60/122)	Reference		
	Yes 27.8 (5/18)	0.3974	0.1335-1.1830	0.0973
Metabolic disorders	No 62/126	Reference		
	Yes 4/14	0.3871	0.1151-1.3016	0.1250

SE = Subclinical endometritis;

RFM = Retained fetal membranes;

CI = Confidence interval.

Cows with SE were less likely to conceive after their first artificial insemination OR = 0.44, $p < 0.05$ than cows without SE (table 4). Also, cows with RFM and metritis had a decreased probability (OR 0.39, $p < 0.05$ and OR 0.33, $p < 0.05$ respectively) to conceive after first artificial insemination.

Cows with SE needed more artificial inseminations to conceive (2.4 ± 0.3) than cows without SE (2.1 ± 0.2). Also, cows with SE had a median of 41 days longer to conceive until 180 DIM compared with cows without SE.

4. Discussion

Our data show that a threshold $\geq 16\%$ was valuable for monitoring the uterine health status during voluntary waiting period (VWP).

Kasimanickam et al., (2004) [19] used a threshold level at $\geq 18\%$ PMN at 20 to 33 days postpartum. Barlund et al., (2008) [21] reported, a threshold at $> 8\%$ PMN at 28 to 41 days pp. In a recent study Lee et al. (2018) [39] reported an optimal threshold $\geq 14\%$ PMN.

There are diverse factors affecting cow's reproductive performance as nutrition, herd health program, cow production, and the climate or barn condition. In general, most authors used PMN % threshold of 15-18% for SE diagnosis at 21-30 days pp, and values of 4-10% for diagnosis at later periods [24].

The prevalence of SE in this study was 34.3%. The reported prevalence of SE varied between 7 and 53% [24].

In this report the retained fetal membranes and metritis were important factors for SE. The OR of 3.020 in this study, for the risk of a RFM was higher than the OR (1.87) reported by Lee et al. (2018) [38]. Gautam et al., (2009) [25] reported on OR of 4024. The OR (4.92) for metritis in this study was higher than the OR (3.07) reported by Lee et al. (2018) [39].

Retained fetal membranes offer a good natrium or bacterial growth. Also, metritis as chronic infection of the uterus causes uterine inflammation. Dystocia and metabolic disorders do not significantly affect for prevalence of SE.

In our study SE significantly affected some subsequent reproductive parameters. Endocrine disturbances in pp period induced a lower probability of cyclicity resolution within 28-30 days in milk.

Ovulation of dominant follicle can be inhibited as a result of bacterial endotoxins that suppress the release of gonadotrophin – releasing hormone and the luteinizing hormone [40].

The impact of milk fever on the health of dairy cows is noticeable. Martinez et al., (2012) [40] observed that cows with serum Ca < 8.59 mg/dl in at least one of the first 3 days pp had reduced neutrophil phagocytic and killing activities in vitro.

Low probability of conception after first AI, more AIs per conception and longer days to conceive until 180 days in milk in cows with SE in this report are comparable to the results of some previous [21, 7, 19, 39].

Prevalence of SE in a dairy farm may reflect the immune status of dairy cows.

4. Conclusions

The cytobrush technic can provide sufficient uterine cells to calculate % PMN threshold. The method is non-invasive and the samples acquired are representative for endometrial cells involved in the uterine immune defense system.

A PMN threshold of 16% was good to define SE at 28-30 days pp, above which there was a significant effect on some reproductive parameters.

Our results can be useful guidance for monitoring uterine health of dairy cows in the pp period.

The control of RFM and metritis after parturition it is required to prevent SE in dairy herd.

Acknowledgements

The research leading to these results has received funding from ARDS Simnic-CRAIOVA. The authors have not stated any conflicts of interest.

References

1. Lazzari G., Duchi R., Colleoni S., Baldazzi L., Benedetti V., Galli A., M. Luini, M. Ferrari, C. Galli. 2011. Le patologie uterine cliniche e subcliniche come causa di infertilità nelle bovine da latte: studio epidemiologico in due allevamenti della regione Lombardia. *Large Animal Review*. 17: 43-47.
2. Madoz L.V., M.J. Giuliadori, A.L. Migliorisi, M. Jaureguiberry, R.L. de la Sota, 2014. Endometrial cytology, biopsy and bacteriology for the diagnosis of subclinical endometritis in grazing dairy cows. *J. of Dairy Sci.* 97(1):195-201. doi: 10.3168/jds.2013-6836.
3. Beatriz Mariño Fuentes, Luis Angel Quintela Arias, Juan José Becerra González, Lidia del Barrio del Sol, Jesús Enrique Mociños Feijóo, José Luis Guillín Puñal, Mónica Barrio López, Alberto Prieto Lago, José Manuel Díaz Cao, Gonzalo Fernández Rodríguez, Pedro José García Herradón, Ana Isabel Peña Martínez. 2017. Agreement between postmortem endometrial cytology, biopsy and bacteriology in culled dairy cow. *Animal Reproduction* 14(4): 1024-1033. <http://dx.doi.org/10.21451/1984-3143-AR826>.
4. Sheldon I.M., Price S.B., Cronin J., Gilbert R.O., Gadsby J.E., 2009. Mechanism of infertility associated

with clinical and subclinical endometritis in high producing dairy cattle. *Reproduction in Domestic Animals*, 44 (suppl. 3) 1-9. doi: 10.1111/j.1439-0531.2009.01465.x.

5. Sheldon I.M., Rycroft A.N., Dogan B., Craven M., Bromfield J.J., Chandler A., Mark H. Roberts, Sian B. Price, Robert O. Gilbert, Kenneth W. Simpson. 2010. Specific strains of *Escherichia coli* are pathogenic for the endometrium of cattle and cause pelvic inflammatory disease in cattle and mice. *PLoS One* 5(2); e9192. doi: 10.1371/journal.pone.0009192.
6. Priest N. Effect of a non-steroidal anti-inflammatory drug on subclinical endometritis in dairy cows and the identification of at – risk cows. (Master Thesis, Department of Agricultural Sciences) New Zealand: Lincoln University; 2013.
7. Gilbert R.O., S.T. Shin, C.L. Guard, H.N. Erb, M. Frajblat, 2005. Prevalence of endometritis and its effects on reproductive performance of dairy cows. *Theriogenology* 64(9): 1879-1888. doi: 10.1016/j.theriogenology.2005.04.022.
8. Dubuc J., Duffield T.F., Leslie K.E., Walton J.S., LeBlanc S.J., 2010. Definition and diagnosis of postpartum endometritis in dairy cows. *Journal of Dairy Science*. 93(11): 5225-5233. doi: 10.3168/jds.2010-3428.
9. Williams E.J., D.P. Fischer, D.E. Noakes, G.C.W. England, A. Rycroft, H. Dobson, I.M. Sheldon. 2007. The relationship between uterine pathogen growth and ovarian function in the postpartum dairy cow. *Theriogenology*. 68(4): 549-559. doi: 10.1016/j.theriogenology.2007.04.056.
10. Lavon Y., Leitner G., Goshen T., Braw-Tal R., Jacoby S., Wolfenson D., 2008. Exposure to endotoxin during estrus alters the timing of ovulation and hormonal concentrations in cows. *Theriogenology* 70(6): 956-967. doi: 10.1016/j.theriogenology.2008.05.058.
11. Herath S., Lilly S.T., Fischer D.P., Williams E.J., Dobson H., Bryant C.E., I.M. Sheldon. 2009. Bacterial lipopolysaccharide induces on endocrine switch from Prostaglandin F2alpha to prostaglandin E2 in bovine endometrium. *Endocrinology*. 150(4): 1912-1920. Doi: 10.1210/en.2008-1379.
12. Shimizu T., Miyauchi K., Shirasuna K, Ballwein H., Magata F., Murayama C., A. Miyamoto. 2012. Effects of lipopolysaccharide (LPS) and peptidoglycan (PGN) on estradiol production in bovine granulosa cells from small and large follicles. *Toxicology In Vitro*.; 26(7): 1134-1142. doi: 10.1016/j.tiv.2012.06.014.
13. Hammon D.S., I.M. Evjen, T.R. Dhiman, J.P. Goff, and J.L. Walters, 2006. Neutrophil function and energy status in Holstein cows with uterine health disorders. *Vet. Immunol. Immunopathol.* 113(1-2): 21-29. doi: 10.1016/j.vetimm.2006.03.022.

14. Hoeben D., E. Monfardini, G. Opsomer, C. Burvenich, H. Dosogne, A. De Kruif, and J.F. Beckers, 2000. Chemiluminescence of bovine polymorphonuclear leucocytes during the periparturient period and relation with metabolic markers and bovine pregnancy – associated glycoprotein. *J. Dairy Res.* 67(2): 249-259. doi: 10.1017/s0022029900004052.
15. Fischer C., M. Drillich, M. Odau, W. Heuwieser, E. Einspanier, C. Gabler, 2010. Selected pro-inflammatory factor transcripts in bovine endometrial epithelial cells are regulated during the oestrous cycle and elevated in case of subclinical or clinical endometritis. *Reproduction, Fertility, and Development.* 22(5): 818-829. doi: 10.1071/RD09120.
16. Gabler C., C. Fischer, M. Drillich, R. Einspanier, W. Heuwieser, 2010. Time-dependent mRNA expression of selected pro-inflammatory factors in the endometrium of primiparous cows postpartum. *Reproductive Biology and Endocrinology.* 8: 152. Doi: 10.1186/1477-7827-8-152.
17. Galvão K.N., N.R. Santos, Galvão J.S., R.O. Gilbert, 2011. Association between endometritis and endometrial cytokine expression in postpartum Holstein cows. *Theriogenology.* 76(2): 290-299. Doi: 10.1016/j.theriogenology.2011.02.006.
18. Ghasemi F., P. Gonzales-Cano, P.J. Griebel, C. Palmer, 2016. Proinflammatory cytokine gene expression in endometrial cytobrush samples harvested from cows with and without subclinical endometritis. *Theriogenology.* 78(7): 1538-1547. Doi:10.1016/j.theriogenology.2012.06.022.
19. Kasimanickam R., T.F. Duffield, R.A. Foster, C.J. Gartley, K.E. Leslie, J.S. Walton, W.H., Johnson, 2004. Endometrial cytology and ultrasonography for the detection of subclinical endometritis in postpartum dairy cows. *Theriogenology.* 62(1-2): 9-23. Doi: 10.1016/j.theriogenology.2003.03.001.
20. Salehi R., M.G. Colazo, M. Gobikrushanth, U. Basu, D.J. Ambrose, 2017. Effects of prepartum oilseed supplements on subclinical endometritis, pro- and anti-inflammatory cytokine transcripts in endometrial cells and postpartum ovarian function in dairy cows. *Reproduction, Fertility, and Development.* 29(4): 747-758. Doi: 10.1071/RD15334.
21. Barlund C.S., T.D. Carruthers, C.L. Waldner, C.W. Palmer, 2008. A comparison of diagnostic techniques for postpartum endometritis in dairy cattle. *Theriogenology.* 69(6): 714-723. Doi: 10.1016/j.theriogenology.2007.12.005.
22. Meira E.B.S. Jr., L.C.S. Henriques, L.R.M. Sá, L. Gregory, 2012. Comparison of ultrasonography and histopathology for the diagnosis of endometritis in Holstein Friesian cows. *J. Dairy Sci.* 95(12): 6969-6973. Doi: 10.3168/jds.2011-4950.
23. Wagener K., C. Gabler, M. Drillich, 2017. A review of the ongoing discussion about definition, diagnosis and pathomechanism of subclinical endometritis in dairy cows. *Theriogenology.* 94: 21-30. doi: 10.1016/j.theriogenology.2017.02.005.
24. Luis Angel Quintela Arias, Marcos Vigo Fernández, Juan José Becerra González, Mónica Barrio López, Pedro José García Herradón and Ana Isabel Peña Martínez, 2018. Subclinical Endometritis in Dairy Cattle. *New Insights into Theriogenology.* Doi: 10.5772/intechopen.80229.
25. Gautam G., Nakao T., Yusuf M., Koike K., 2009. Prevalence of endometritis during the postpartum period and its impact on subsequent reproduction in two Japanese dairy herd. *Animal Reproduction Science.* 116(3-4): 175-187. doi: 10.1016/j.anireprosci.2009.02.001.
26. Burke C.R., Meire S., McDougall S., Compton C., Mitchell M., Roche J.R., 2010. Relationship between endometritis and metabolic state during the transition period in pasture-grazed dairy cows. *J. Dairy Science.* 93(11): 5363-5373. doi: 10.3168/jds.2010-3356.
27. Green M.P., Ledgard A.M., Berg M.C., Peterson A.J., Back P.J., 2009. Prevalence and identification of systemic markers of sub-clinical endometritis in postpartum dairy cows. *Proceeding of the New Zealand Society of Animal Production.* 69: 37-42.
28. LeBlanc S.J., Duffield T.F., Leslie K.E., Bateman K.G., Keefe G.P., Walton J.S., Johnson W.H. 2002. The effect of treatment of clinical endometritis on reproductive performance in dairy cows. *J. Dairy Sci.* 85(9): 2237-2249. doi: 10.3168/jds.S0022-0302(02)74303-8.
29. Bretzlaff K., 1987. Rationale for treatment of endometritis in the dairy cow. *Vet. Clin. North Am. Food Anim. Pract.* 3(3):593-607. doi: 10.1016/s0749-0720(15)31132-4.
30. Pérez-Marin C.C. and L.A. Quintela. 2023. Current insights in the Repeat Breeder Cow Syndrome. *Animals* 13(13): 2187. Doi.org/10.3390/ani.13132187.
31. Prunner I., Wagener K., Pothmann H., Ehling-Schulz M., Drillich M. 2014. Risk factors for uterine diseases on small- and medium- sized dairy farm determined by clinical, bacteriological, and cytological examinations. *Theriogenology.* 82(6): 857-865. Doi:10.1016/j.theriogenology.2014.06.015.
32. Gobikrushanth M., Salehi R., Ambrose D.J., Colazo M.G., 2016. Categorization of endometritis and its association with ovarian follicular growth and ovulation, reproductive performance, dry matter intake and milk yield in dairy cattle. *Theriogenology* 86(7): 1842-1849. Doi:10.1016/j.theriogenology.2016.06.003.
33. Green M.P., Ledgard A.M., Beaumont S.E., Berg M.C., McNatty K.P., Peterson A.J., P J Back. 2011. Long-term alteration of follicular steroid concentrations in relation to subclinical endometritis in postpartum dairy cows. *Journal of Animal Science.* 89(11): 3551-3560. Doi:10.2527/jas.2011-3958.
34. Fernandez-Sanchez F.I., Barrio-Lopez M., Quintela-Arias L.A., Becerra-Gonzales J.J., Peña-

- Martinez A.I., Martinez-Bello D., Pedro J. Garcia-Herradon and Carlos C. Perez-Marín. 2014. Use of endometrial cytology and metabolic profiles for selection of embryo donor cows. *Spanish Journal of Agriculture Research*. 12(3): 664-671. Doi: 10.5424/sjar/2014123-4948.
35. Barrio M., Becerra J.J., Herradon P.G., Sebastian F., Fernandez M., Quintela L.A., Vigo M. et al., 2015. Influence of subclinical endometritis on the reproductive performance of dairy cows. *Spanish Journal of Agricultural Research*. 13(4): 05-02. Doi 10.5424/sjar/2015134-8129.
36. Bell M.J., Roberts D.J. 2007. The impact of uterine injection on a dairy cow's performance. *Theriogenology*. 68(7): 1074-1079. Doi: 10.1016/j.theriogenology.2007.08.010.
37. Kasimanickam R., T.F. Duffield, R.A., Foster, K.J. Gartley, K.E. Leslie, J.S. Walton, and W.H. Johnson. 2005. A comparison of the cytobrush and uterine lavage techniques to evaluate endometrial cytology in clinically normal postpartum dairy cows. *Can. Vet. J.* 46(3): 255-259.
38. Lombard J.E., Garry F.B., Tomlinson S.M., Garber L.P. 2007. Impacts of dystocia on health and survival of dairy calves. *J. Dairy Sci.* 90(4): 1751-1760. Doi: 10.3168/jds.2006-295.
39. Lee S.C., Jeong J.K., I.S. Choi, H.G. Kang, Y.H. Jung, S.B. Park, I.H. Kim. 2018. Cytological endometritis in dairy cows: diagnostic threshold, risk factors and impact on reproductive performance. *J. Vet. Sci.* 19(2): 301-308. Doi: 10.4142/jvs.2018.19.2.301.
40. Martinez N., Risco C.A., Lima F.S., Bisinotto R.S., Greco L.F., Ribeiro E.S., Maunsell F., Galvão K.N., Santos J.E.P. 2012. Evaluation of peripartal calcium status, energetic profile and neutrophil function in dairy cows at low or high risk of developing uterine disease. 95(12):7158-72. doi: 10.3168/jds.2012-5812.